ENCEPHALITIS / INITIAL CASE REPORT FORM

PATIENT INFORMATION					
Last Name		ame		County	
Address		City		Zipcode	State
Telephone-H () Occupation:	W ()		Date of Birth	//	Age
Occupation:	Race:	□White	□Black □Am	Indian/Alaskan	☐ Asian ☐ Other
Ethnicity: ☐ Hispanic ☐ Non-Hispa	nic Unknown	Sex:	☐ Male ☐ Female	Pregnant:	s 🗆 No 🗆 Unknown
CLINICAL INFORMATION					
Street Address	spitai Naine		ity	State	 7in
Hospitalized? Hospitalized? Hospitalized. Hospitalized? Hospitalized. H	Date of a	dmission	/ / Da	ate of discharge/tran	sfer / /
Date of first symptoms//	Date of fir	st neurolog	gic symptoms/_	/	~~~~
Current Diagnosis.	nus 🗀 meningo	енсернані	is 🗀 meningius	□ 0tilei	
Initial Diagnosis:	litis 🛮 meningo	encephalit	is	other	
Fever (\geq 38EC or 100EF) \square Yes \square Headache \square Yes \square	INo ⊔Unknow INo □Unknow	n	Altered mental stat	us	□ No □ Unknown □ No □ Unknown
	No ☐ Unknow	II n	Muscle weakness	ai sigiis ⊔ 1es □ Ves	□ No □ Unknown
	No Unknow	n	Previous Flavirus v	accination \(\bar{\pi} \) Yes	□ No □ Unknown
Other neurologic signs	No ☐ Unknow	n If yes	s, describe		
Other symptoms (current or 1 month	before onset)				
Outcome Recovered Died	□Unknown	If patient	died, date of death _	/	
LABORATORY INFORMATION	/ TEST RESULT	<u>'S</u>	_		
CSF (specify units) Date/_ Glu Prot Gram stain CBC (specify units) Date/_	/ Ab	onormal?	☐ Yes ☐ No ☐	Unknown	o/ T 1
Glu Prot	RBC	W	/BC	Diff: Segs	% Lymphs
CRC (specify units) Date /		Culture ₋	Diff: Sage	% I vmph	p 0/ ₆
CBC (specify units) Date/_ MRI Date// CT Date/ EEG Date/	Results W	вс	Dill. Segs _	% Lympn	S70
CT Date/	Results				
EEG Date/	Results				
Microbiology / serology	Results				
CURRENT TREATMENT T	ype:			Date	e started:
					/ /
(and that of antibacterial)					
,					
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Division of Consolidated Laboratory Services (DCLS) - Arbovirus Testing

Testing Criteria for Encephalitis: Any adult or pediatric patient admitted to a hospital with a presumed diagnosis of viral encephalitis, or with focal CNS findings and fever should submit a whole blood or serum sample for diagnostic testing at DCLS following the guidelines listed below. **Recommended Criteria for Suspect Cases of WNV** - Any patient with viral encephalitis (Criteria a, b and c below) with or without associated muscle weakness (Criteria d)

- a. Fever $> 38^{\circ}$ C or 100° F, and
- b. Altered mental status (altered level of consciousness, agitation, lethargy) and/or other evidence of cortical involvement (e.g., focal neurologic findings, seizures), and
- c. CSF pleocytosis with predominant lymphocytes and/or elevated protein and a negative gram stain and culture, and/or
- d. Muscle weakness (especially flaccid paralysis) confirmed by neurologic exam or by EMG.

Testing Systems Employed: Serological specimens submitted for arbovirus testing will be evaluated using an IgM Antibody Capture Enzyme-linked Immunosorbent Assay (MAC-ELISA) and an IgG ELISA to identify St. Louis Encephalitis (SLE), Eastern Equine Encephalitis (EEE), LaCrosse Encephalitis (LAC), and WNV -reactive antibody. If SLE, EEE, LAC, or WNV-reactive antibodies are present, specimens will be forwarded to CDC for Plaque Reduction Neutralization Testing (PRNT) confirmation. Real-time RT-PCR will be performed on biopsy and postmortem tissue specimens, and acute serum and CSF. Viral isolation is available upon request.

Specimen Types and Amounts:

Specimen	Test	Specimen volume	Shipment
Paired acute-phase (0-8 days post-onset of illness) and	MAC-ELISA	2 ml sera in plastic tube (preferred)	Refrigerated
convalescent-phase (14-21 days after acute specimen) sera	IgG ELISA	or 5 ml whole coagulated blood	
Acute-phase CSF	MAC-ELISA	1.0 ml in plastic tube	Frozen
	Viral Isolation		
	Real-time RT-PCR		
Brain, biopsy or postmortem tissue	Viral Isolation	1 gram	Frozen
	Real-time RT-PCR		

Submission Form: Include a Encephalitis/Initial Case Report Form for each patient to be tested. Fill out the form as completely as possible. Accurate interpretation of serologic findings requires knowledge of the specimen. It is imperative that the following data accompany specimens submitted: 1) symptom onset date; 2) date of sample collection; 3) unusual immunological status of patient; 4) current address and travel history; 5) history of prior vaccination against flavivirus disease; and 6) brief clinical summary including suspected diagnosis.

Packaging for Shipment:

Serum or whole coagulated blood samples: If you are requesting serology, the specimen should be kept cool but there is no need to keep it frozen. Wrap your specimen in absorbent material, pack it in a sealable plastic bag, place the bag in a rigid container (styrofoam box or cardboard mailing container), place the container and a cold pack in a styrofoam cooler for shipment.

CSF or tissue: If you are requesting virus isolation or Real-time RT-PCR for WNV on tissue or CSF, the specimen should be frozen in a sterile container prior to shipment and it should be sent on enough dry ice to insure it will remain frozen until receipt. These specimens should be wrapped in absorbent material, packed in a sealable plastic bag, placed in a second sealable bag, and then placed in a cooler with dry ice for shipment.

Coolers should be shipped 24 hour/overnight delivery.

If you are hand carrying the specimen, please observe the packing instructions above to secure the specimen in transit.

Sample Shipment:

Non-courier shipment:

Package specimens as indicated above and send to DCLS using the following address:

Division of Consolidated Laboratory Services

1 North 14th Street, SRM Room 151

Richmond, Virginia 23219-3691

Courier shipment:

Package specimens as indicated above and send via DCLS courier.

Reporting of Test Results: Test results are normally available 10 days to 2 weeks after receipt of specimens. During periods of heavy submission, turn around times may be longer. Receipt of a hard copy of the results may take 2 weeks after completion of testing. If initial screening tests are positive, results will be communicated to Dr. Suzanne Jenkins prior to the availability of confirmatory test results. Confirmatory testing performed on screen positive samples will delay the reporting of results to the submitter.

Please contact Denise Pettit (804-786-9715) at DCLS if you have questions regarding sample collection or shipment.